Analysis of the intraspinal calcium dynamics and its implications for the plasticity of spiking neurons

Luk C. Yeung

Physics Department and Institute for Brain and Neural Systems, Brown University, Box 1843, Providence, Rhode Island 02912, USA

Gastone C. Castellani

Dipartimento Fisica, Università Bologna, Viale Berti Pichat 6/2, Bologna 40127, Italy

Harel Z. Shouval*

Physics Department, Brown University, Box 1843, Providence, Rhode Island 02912, USA (Received 21 April 2003; revised manuscript received 25 June 2003; published 26 January 2004)

The influx of calcium ions into the dendritic spines through the N-methyl-D-aspartate (NMDA) channels is believed to be the primary trigger for various forms of synaptic plasticity. In this paper, the authors calculate analytically the mean values of the calcium transients elicited by a spiking neuron undergoing a simple model of ionic currents and back-propagating action potentials. The relative variability of these transients, due to the stochastic nature of synaptic transmission, is further considered using a simple Markov model of NMDA receptors. One finds that both the mean value and the variability depend on the timing between presynaptic and postsynaptic action potentials. These results could have implications for the expected form of the synaptic-plasticity curve and can form a basis for a unified theory of spike-time-dependent, and rate-based plasticity.

DOI: 10.1103/PhysRevE.69.011907

PACS number(s): 87.16.Ac, 87.16.Uv, 87.17.Aa, 87.17.Nn

I. INTRODUCTION

Calcium ions are ubiquitous mediators of metabolic change in many cellular systems. In the cortex, they are believed to be the primary chemical signal for the induction of bidirectional synaptic plasticity [1-3], which is widely accepted to be the basis for many forms of learning, memory, and development.

Experimentally, bidirectional synaptic plasticity can be induced by various different induction protocols [4-6], including the recently characterized spike-time-dependent plasticity (STDP) [7,8]. In this induction method, if a presynaptic action potential occurs within a window of a few tens of milliseconds before a postsynaptic action potential (Δt >0), long-term potentiation (LTP) is elicited; if the order is switched ($\Delta t < 0$), long-term depression (LTD) happens. Interestingly. STDP and many other induction protocols share a common property, which is their dependence on the amount of integrated activation of the N-methyl-D-aspartate receptors (NMDAR) during conditioning. NMDAR are natural coincidence detectors; their activation relies both on the efficiency of neurotransmitter release from the presynaptic cell and on the level of depolarization of the postsynaptic cell. Most remarkably, they are permeable to calcium ions, which suggests a functional link between calcium influx through NMDAR and the induction of LTP and LTD.

Recently, several dynamical models have been proposed to account for STDP [9-12], the relationship between STDP and rate-based models has been examined using averaging methods [13,14], and there have been attempts to account for

receptive field formation on the basis of STDP [15]. We have recently proposed a unified theory of synaptic plasticity [16] based on the following assumptions: (1) the level of intracellular calcium concentration determines the sign and magnitude of synaptic plasticity [17-19]-when calcium falls below a threshold θ_d , no plasticity occurs, when it falls between θ_d and θ_p , $\theta_p > \theta_d$, LTD is induced, and for calcium above θ_n , LTP is induced; (2) the relevant sources of calcium are the NMDA channels; and (3) dendritic backpropagating action potentials (BPAPs) contributing to STDP have a slow "after-depolarizing" tail component. We have shown that these simple assumptions are sufficient to account for the various experimental plasticity-induction protocols. In addition, this model has also produced previously uncharacterized predictions, such as: (a) the shape of the STDP learning curve should be frequency-dependent and (b) there should be a novel form of spike time-dependent LTD for Δt larger than the LTP-inducing intervals. Recent experimental results are consistent with these predictions [20,21].

In this paper, we present a systematic calculation of the calcium transients as a function of prespike and postspike timings, as well as of the neuronal firing rates. In Sec. II we derive the solution for the mean calcium transients in the simplest case where the BPAP consists of a single exponential, and show that its dependence on Δt is rather intuitive. A more realistic approach is taken in Sec. III, where the BPAP is composed of a sum of two exponentials with different time constants. This more complex assumption is in closer agreement with our previous work [16] and is consistent with experimental observations [22]. The analysis shows that the slow component contributes for different calcium levels in the baseline condition (pre only) and the post-pre condition $(\Delta t < 0)$, while the fast component sharpens the difference of the calcium levels in the transition between post/pre (Δt <0) and pre/post ($\Delta t > 0$) conditions. The sharpness of this

^{*}Present address: Department of Neurobiology and Anatomy, The University of Texas Medical School at Houston, Houston, TX 77030. Electronic address: harel.shouval@uth.tmc.edu

transition is shown to be further enhanced by the appropriate choice of the calcium decay time constant. For completeness, in Sec. IV, we study the effects of calcium accumulation due to repetitive firing which affects the rate-dependence of some plasticity-induction protocols including the STDP learning window. Finally, in Sec. V we take into account the stochastic properties of synaptic transmission and estimate the trialby-trial variability of the calcium transients. This variability can be significant if the number of postsynaptic NMDAR in each spine is small. Our analysis demonstrates that this variability depends on Δt , increasing with Δt for $\Delta t > 0$. These results could significantly alter the form of the STDP curve under different experimental conditions.

II. ANALYSIS OF CALCIUM INFLUX THROUGH NMDAR

The postsynaptic calcium ions are removed from the intraspinal compartments both through diffusion and through binding to calcium buffers. Let these phenomena be characterized by a single time constant τ , and assume that the calcium dynamics follow a first-order linear differential equation,

$$\frac{d[\operatorname{Ca}]}{dt} = I(t) - \frac{1}{\tau}[\operatorname{Ca}], \qquad (1)$$

where [Ca] is the intracellular calcium concentration, I(t) is the inward calcium current, and τ is the time constant of the passive decay process. This simple dynamics is to the first approximation consistent with experiments [23,24]. Given a initial condition [Ca](0), Eq. (1) has a solution of the form

$$[Ca](t) = e^{-t/\tau} \left[\int_0^t e^{t/\tau} I(t') dt' + [Ca](0) \right].$$
(2)

Note that if the calcium current is a superposition of separate current sources, the calcium concentration can also be decomposed into such a sum.

In this paper we consider the NMDA channels as the sole source of calcium currents. These channels are voltagedependent [25], and have a long open time that can usually be described as a sum of two exponentials [26], thus their currents can be generally described as

$$I(t, \{t^{pre}\}, \{t^{post}\}) = \overline{g}\mathcal{H}(V)f, \qquad (3)$$

where $\{t^{pre}\}\$ and $\{t^{post}\}\$ are the sets of times of presynaptic and postsynaptic action potentials (APs) that occurred prior to time t and \overline{g} is the mean total conductance of the population of NMDAR in a synapse. The \mathcal{H} -function represents the voltage-dependence of calcium influx through NMDAR, and f is the fraction of NMDAR in the open state. It is easy to see that $V = V(t, \{t^{pre}\}, \{t^{post}\})$ and $f = f(t, \{t^{pre}\})$, where the time-dependence of f is a double exponential.

In this section, however, we will focus on the simple scenario of a single pair of presynaptic and postsynaptic APs, where the dynamics of the NMDAR and the BPAP are described as single exponentials with time constants τ_N and τ_B , respectively. We will leave the more realistic two-component-BPAP case to the following section. Unless oth-

erwise mentioned, we replace the stochastic variable f by its mean without change of notation. Thus, for a single presynaptic spike at time t^{pre} , we can write

$$f(t,t^{pre}) = \mu \Theta(t-t^{pre})e^{-(t-t^{pre})/\tau_{\rm N}},\tag{4}$$

where Θ is the Heaviside function and μ is the probability that a channel will open given a presynaptic AP. Let us assume that the postsynaptic depolarization is dominated by the BPAP. Therefore, V in Eq. (3) can be written as

$$V(t, t^{post}) = V_R + V_B(t) \tag{5}$$

$$= V_R + v \Theta(t - t^{post}) e^{-(t - t^{post})/\tau_{\rm B}},$$
(6)

where V_R is resting membrane potential, and $V_B(t)$ describes the dynamics of a BPAP with magnitude v due to a postsynaptic AP occurring at time t^{post} . In general, \mathcal{H} is a nonlinear and nonmonotonic function of V. For tractability, we will make a major simplification by linearizing the \mathcal{H} function in the interval $[V_R, V_R + v]$,

$$\mathcal{H}(V) = a + bV. \tag{7}$$

The original \mathcal{H} function used and its linear fit are displayed in Fig. 10. In Appendix B, we assess the validity of this approximation. By using Eqs. (4) through (7), Eq. (3) becomes

$$I(t, t^{pre}, t^{post}) = \overline{g}(a + bV_R + bv\Theta(t - t^{post}))$$

$$\times e^{-(t - t_{post})/\tau_{\rm B}} \mu\Theta(t - t^{pre})e^{-(t - t^{pre})/\tau_{\rm N}}$$

$$= \overline{g}\mu(a + bV_R)\Theta(t - t^{pre})e^{-(t - t^{pre})/\tau_{\rm N}}$$

$$+ \overline{g}\mu bv\Theta(t - t^{post})\Theta(t - t^{pre})$$

$$+ e^{-(t - t^{pre})/\tau_{\rm N}}e^{-(t - t^{post})/\tau_{\rm B}}$$

$$= I^{pre}(t) + I^{+/-}(t). \qquad (8)$$

The current separates into two components, one that depends only on the timing of the presynaptic AP (I^{pre}) and a second, associative term that depends on the relative timing between the presynaptic and postsynaptic APs $(I^{+/-})$. The associative term will give rise to the differences between pre/post and post/pre conditions, which is essential to STDP learning.

Let $\Delta t = t^{post} - t^{pre}$. It is straightforward to show that

$$I^{+/-} = \begin{cases} I_{peak}^{+} \Theta(t-t^{post}) e^{-(t-t^{post})/\tau_{1}} & \text{if } \Delta t > 0\\ I_{peak}^{-} \Theta(t-t^{pre}) e^{-(t-t^{pre})/\tau_{1}} & \text{otherwise,} \end{cases}$$
(9)

where $\tau_1^{-1} = \tau_B^{-1} + \tau_N^{-1}$, and $I_{peak}^+ = I_{peak}^+(\Delta t)$ and $I^- = I^-(\Delta t)$ denote the peak magnitudes of the pre/post component of the calcium current for $\Delta t > 0$ and $\Delta t < 0$, respectively:

$$I_{peak}^{+}(\Delta t) = \overline{g} \,\mu b v \, e^{-\Delta t/\tau_{\rm N}},\tag{10a}$$

$$I_{peak}^{-}(\Delta t) = \bar{g}\,\mu b v e^{\Delta t/\tau_{\rm B}}.$$
(10b)



FIG. 1. Associative component of the peak calcium current as a function of Δt . $\overline{g}b = 1.5 \times 10^{-3}$, $\mu = 0.8$, v = 60 mV, $\tau_{\rm N} = 100$ ms, and $\tau_{\rm B} = 20$ ms.

The dependence of the peak current on Δt is quite simple (Fig. 1). For $\Delta t > 0$ it decays exponentially with the time constant of the NMDAR open probability τ_N ; whereas for $\Delta t < 0$ it decays with τ_B , the width of $V_B(t)$. It is straightforward to see that, if $\Delta t < 0$ yields calcium levels corresponding to LTD, and if to each calcium concentration is associated a degree of plasticity, there should be a $\Delta t > 0$ region that also corresponds to LTD. The difference is that, since $\tau_B \ll \tau_N$, θ_d in the post/pre region is reached for values of Δt much closer to zero than it is for the pre/post region. Such difference will be even greater in the case where the dynamics of the BPAP is described by two time constants instead of one (see Sec. III).

Using Eqs. (8) and (9), we can rewrite Eq. (2) in terms of each of the components of the calcium current,

$$[Ca](t) = [Ca]^{pre}(t) + \Theta(\Delta t)[Ca]^{+}(t)$$
$$+ \Theta(-\Delta t)[Ca]^{-}(t) + [Ca](0)e^{-t/\tau}, \quad (11)$$

with

$$[Ca]^{pre}(t) = e^{-t/\tau} \int I^{pre}(t') e^{t'/\tau} dt', \qquad (12a)$$

$$[Ca]^{+}(t) = e^{-t/\tau} \int I^{+}(t') e^{t'/\tau} dt', \qquad (12b)$$

$$[Ca]^{-}(t) = e^{-t/\tau} \int I^{-}(t') e^{t'/\tau} dt'.$$
 (12c)



Integrating explicitly, we have

$$[Ca]^{pre}(t) = \overline{g} \mu (a + bV_R) \tau_2 \Theta (t - t^{pre}) (e^{-(t - t^{pre})/\tau_N} - e^{-(t - t^{pre})/\tau}), \qquad (12a')$$

$$[Ca]^{+}(t) = I_{peak}^{+} \tau_{3} \Theta(t - t^{post}) (e^{-(t - t^{post})/\tau_{1}} - e^{-(t - t^{post})/\tau}),$$
(12b')

$$[Ca]^{-}(t) = I_{peak}^{-} \tau_3 \Theta(t - t^{pre}) (e^{-(t - t^{pre})/\tau_1} - e^{-(t - t^{pre})/\tau}),$$
(12c')

where $\tau_2^{-1} = \tau^{-1} - \tau_N^{-1}$ and $\tau_3^{-1} = \tau^{-1} - \tau_N^{-1} - \tau_B^{-1}$. Examples of calcium transients are displayed in Fig. 2,

Examples of calcium transients are displayed in Fig. 2, where $[Ca]^{pre/post} = [Ca]^{pre} + [Ca]^+$ and $[Ca]^{post/pre} = [Ca]^{pre} + [Ca]^-$. Naturally, the $[Ca]^{pre}$ term is identical for the pre-post [Fig. 2(a), $\Delta t = 10$ ms] and the post-pre [Fig. 2(b), $\Delta t = -10$ ms] conditions (dashed line). The difference between these conditions lies in the associative term (dashdotted line). $[Ca]^+$ raises at the point $t = t^{post}$ and $[Ca]^$ raises at $t = t^{pre}$, their relative amplitudes being determined by $I_{peak}^{+/-}$ (Fig. 1).

III. A BACK-PROPAGATING ACTION POTENTIAL WITH TWO COMPONENTS

The model described in the preceding section assumes that $V_B(t)$ decays exponentially with a single time constant. However, there are experimental indications that an additional slow after-depolarizing component exists in some cell bodies [27] and dendrites [22,28]. The dynamics of the BPAP defines to what extent the postsynaptic spike-timing interacts with the presynaptic one; in particular, a slower decaying process ensures that such interaction spans for a period of tens of milliseconds, as required by the STDP learning rule. In this section, we consider that $V_B(t)$ is composed by a sum of two exponentials, with a slow τ_B^s and a fast τ_B^f time constants, respectively. Equation (6) should therefore be rewritten as

$$V(t,t^{post}) = V_R + v \Theta(t-t^{post})$$
$$\times [v^f e^{-(t-t^{post})/\tau_{\rm B}^f} + v^s e^{-(t-t^{post})/\tau_{\rm B}^s}], \quad (6')$$

where v^s , v^f , τ^s_B , and τ^f_B are the relative amplitudes and time constants of the slow and fast components of $V_B(t)$, and

FIG. 2. The dynamics of the total calcium (solid line) is composed of two different contributions: the calcium due to the prespike alone (dashed line) and the calcium due to the association of prespike and postspike (dash-dotted line). $a\overline{g} = 1.03 \times 10^{-1}$, $V_R = -65$ mV, $\tau = 50$ ms, [Ca](0)=0 and the onset of t_{pre} is at time zero. The remaining parameters are as in Fig. 1. (a) $\Delta t = 10$ ms, (b) $\Delta t = -10$ ms.



FIG. 3. (a) Associative component of the peak calcium current as a function of Δt , for $v^f = 0.75$, $v^s = 0.25$, $\tau_B^f = 3$ ms, and $\tau_B^s = 35$ ms. The dotted line shows the previous, single-component BPAP result, for comparison. (b) Total calcium transients for $\Delta t = 10$ ms (dashed line), -10 ms (dash-dotted line) and [Ca]^{*pre*} (solid line). All the other parameters are as in Figs. 1 and 2.

respectively, $(v^s, v^f) > 0, v^s + v^f = 1$. Analogously to the preceding section, the calcium currents from Eq. (9) can be separated into two components, I^{pre} and $I^{+/-}$. I^{pre} is identical to what we have derived before, whereas $I^{+/-}$ reads

$$I^{+/-}(t) = \begin{cases} \Theta(t - t^{post}) I_{peak}^{+} [v^{f} e^{-(t - t^{post})/\tau_{1}^{f}} + v^{s} e^{-(t - t^{post})/\tau_{1}^{s}}] & \text{if } \Delta t > 0 \\ \Theta(t - t^{pre}) [I_{peak}^{-f} v^{f} e^{-(t - t^{pre})/\tau_{1}^{f}} + I_{peak}^{-s} v^{s} e^{-(t - t^{pre})/\tau_{1}^{s}}] & \text{otherwise,} \end{cases}$$
(9')

where I_{peak}^+ is as defined in Eq. (10a), $I_{peak}^{-f} = \overline{g} \mu b v e^{\Delta t / \tau_{\rm B}^{J}}$, $I_{peak}^{-s} = \overline{g} \mu b v e^{\Delta t / \tau_{\rm B}^{S}}$. The calcium current $I^{pre/post}$ peaks at $t = t^{post}$ for $\Delta t > 0$ with amplitude I_{peak}^+ and at $t = t^{pre}$ for $\Delta t < 0$ with amplitude $I_{peak}^- = v^s I_{peak}^{-s} + v^f I_{peak}^{-f}$. Thus, I_{peak}^- decays more abruptly than its single-component-BPAP counterpart [Fig. 3(a)].

We can again integrate each component of the calcium currents separately.

$$[Ca]^{pre}(t) = \overline{g} \mu (a + bV_R) \tau_2 \Theta (t - t^{pre}) \\ \times (e^{-(t - t^{pre})/\tau_N} - e^{-(t - t^{pre})/\tau}), \quad (12a'') \\ [Ca]^+(t) = I^+_{peak} \Theta (t - t^{post}) \{ v^f \tau_3^f (e^{-(t - t^{post})/\tau_1^f} \\ - e^{-(t - t^{post})/\tau}) + v^s \tau_3^s (e^{-(t - t^{post})/\tau_1^s} \\ - e^{-(t - t^{post})/\tau}) \}, \quad (12b'')$$

$$[Ca]^{-}(t) = \Theta(t - t^{pre}) \{ I_{peak}^{-f} v^{f} \tau_{3}^{f} (e^{-(t - t^{pre})/\tau_{1}^{f}} - e^{-(t - t^{post})/\tau}) \}$$

$$+I^{-s}v^{s}\tau_{3}^{s}(e^{-(t-t^{pre})/\tau_{1}^{s}}-e^{-(t-t^{pre})/\tau})\}, \quad (12c'')$$



where $(\tau_1^f)^{-1} = (\tau_B^f)^{-1} + \tau_N^{-1}$, $(\tau_3^f)^{-1} = \tau^{-1} - \tau_N^{-1} - (\tau_B^f)^{-1}$, and τ_1^s and τ_3^s are defined similarly to τ_B^s , but with τ_B^s replacing τ_B^f .

The effect of the slow component of the function $V_B(t)$ on the calcium transients is shown in Figs. 3(b) and 4(a). Since both $[Ca]^+$ and $[Ca]^-$ have the same linear dependence on the magnitude v_s of this slow component, it primarily contributes to the separation between the pre-only condition (diamond), and the associative conditions (circle and square).

A longer decaying tail for the postsynaptic variable provides a longer time interval in which interaction with the presynaptic variables is possible. Thus, the dynamics of the presynaptic variable, such as the calcium time constant τ , will also influence the relative magnitudes of the calcium transients for the pre-post and post-pre conditions. Indeed, the combination of a faster calcium kinetics with the different BPAP time courses enhances the difference between these magnitudes [Fig. 4(b)]. Therefore, a two-component BPAP model produces a sharper transition of the learning curve between the post-pre LTD and the pre-post LTP. Initial estimates of the calcium time constant in the spines were of

> FIG. 4. (a) Peak calcium transients as a function of the slow component of the V_B function v^s for the pre-before-post (circle), post-before-pre (square), and the pre-only (diamond) conditions. (b) Relative magnitudes of the peak calcium transients, between pre and post-pre (circle), and post-pre and pre-post (square) conditions, as a function of the calcium time constant τ . All parameters are as in Fig. 3.

011907-4

the order of 70–125 ms [23,29]. However, these values are obtained through calcium imaging, whose results may depend on the kinetics of the calcium indicators used, limiting the accuracy of the estimates. Recent experimental techniques, in which the effect of the calcium buffers are minimized, have lead to time constants of as low as 20 ms [24].

IV. RATE-DEPENDENCE OF THE CALCIUM TRANSIENTS

In the previous sections we have analyzed the calcium transients evoked by one pair of presynaptic and postsynaptic spikes, these have significant implications for STDP at low frequency. However, STDP at low frequency is not the sole method for inducing synaptic plasticity, and in some cases even STDP requires that the pairs of prespike and postspike are delivered above a certain frequency [7,21]. Thus it is important to analyze the contribution of spike delivery frequency ν , to the calcium concentration, in addition to the relative timing effect. We analyze the case in which both presynaptic and postsynaptic neurons fire regularly at the same frequency ν , with the onset of the postsynaptic spike-train shifted from that of the presynaptic spike-train by $\Delta t < 1/\nu$. Note, however, that in the most general case the number of presynaptic and postsynaptic spikes as well as their timing differences will vary.

Recall that the NMDA current in Eq. (3) is the product between the fraction of opened channels f(t) and the linearized Mg-block function $\mathcal{H}(V(t)) = a + bV(t)$, where V(t) $= V_R + V_B(t)$ omits depolarization due to the excitatory postsynaptic potentials. For a sequence $\{t_1^{pre}, t_2^{pre}, \ldots, t_M^{pre}\}$ of presynaptic spike times and $\{t_1^{post}, t_2^{post}, \ldots, t_M^{post}\}$ of postsynaptic spike times, we can write

$$f(t) = \sum_{n=1}^{N} A_n \exp\left(-\frac{t - t_n^{pre}}{\tau_N}\right) \Theta(t - t_n^{pre}), \qquad (13)$$

$$V_B(t) = v \sum_{m=1}^{M-1} \left[\exp\left(-\frac{t - t_m^{post}}{\tau_B}\right) \Theta(t - t_m^{post}) \Theta(-t + t_{m+1}^{post}) \right] + \exp\left(-\frac{t - t_M^{post}}{\tau_B}\right) \Theta(t - t_M^{post}),$$
(14)

where *N* and *M* are, respectively, the total number of prespikes and postspikes before time *t*, and A_n is the increase of the fraction of opened channels upon each prespike *n*. Let f(t) be at a level B_n immediately before the prespike *n* occurs. If the NMDA opening is proportional to the amount of nonopened channels, then $A_n = \mu(1-B_n)$, where μ is the fraction of previously closed channels that open due to the presynaptic spike, $\mu < 1$. Writing B_n explicitly for each *n*, it is easy to see that it satisfies the following expression:

$$B_n = \mu \sum_{i=1}^{n-1} e^{-i/\nu \tau_{\rm N}} = \mu e^{-1/\nu \tau_{\rm N}} \frac{1 - (1 - \mu)^n e^{-n/\nu \tau_{\rm N}}}{1 - (1 - \mu) e^{-1/\nu \tau_{\rm N}}}$$
(15)

$$B_{\infty} = \lim_{n \to \infty} B_n = \frac{\mu e^{-1/\nu \tau_{\rm N}}}{1 - (1 - \mu)e^{-1/\nu \tau_{\rm N}}}.$$
 (16)

With this, Eq. (13) can be written as

$$f(t) = \sum_{n=1}^{N-1} (B_n + A_n) \exp\left(-\frac{t - t_n^{pre}}{\tau_N}\right) \Theta(t - t_n^{pre})$$

$$\times \Theta(-t + t_{n+1}^{pre}) + (A_N + B_N)$$

$$\times \exp\left(-\frac{t - t_N^{pre}}{\tau_N}\right) \Theta(t - t_N^{pre})$$

$$= \sum_n^{N-1} [B_n(1 - \mu) + \mu] \exp\left(-\frac{t - t_n^{pre}}{\tau_N}\right) \Theta(t - t_n^{pre})$$

$$\times \Theta(-t + t_{n+1}^{pre}) [B_N(1 - \mu) + \mu]$$

$$\times \exp\left(-\frac{t - t_N^{pre}}{\tau_N}\right) \Theta(t - t_N^{pre}). \quad (17)$$

We can define, analogously to Eq. (8) and the expressions (12a) through (12c) from Sec. II, the following variables: $I^{pre} = \overline{g}(a+bV_R)f(t), I^{+/-} = \overline{g}bV_B(t)f(t)$, and

$$[Ca]^{pre}(t) = e^{-t/\tau} \int I^{pre}(t') e^{t'/\tau} dt', \qquad (18a)$$

$$[Ca]^{+/-}(t) = e^{-t/\tau} \int I^{pre/post}(t') e^{t'/\tau} dt'.$$
 (18b)

Using Eq. (17), Eq. (18a) reads

$$[Ca]^{pre} = \overline{g} e^{-t/\tau} (a+bV_R) \tau_2 \times \left\{ (e^{1/\nu\tau_2} - 1) \sum_{n=1}^{N-1} [B_n(1-\mu) + \mu] e^{t_n^{pre}/\tau} + (e^{t/\tau_2} - e^{t_N^{pre}/\tau_2}) [B_N(1-\mu) + \mu] e^{t_N^{pre}/\tau_N} \right\}.$$
(19)

We use the approximation $B_n = B_\infty$, and write the time indexes with respect to the timing of the first spike,

$$t_n^{pre} = t_1^{pre} + \frac{n-1}{\nu},$$
 (20a)

$$t_N^{pre} = t_1^{pre} + \frac{\lfloor \nu t \rfloor - 1}{\nu}, \quad \text{and}$$
 (20b)

$$N = \lfloor (t - t_1^{pre})\nu \rfloor + 1, \qquad (20c)$$

where | | is the floor operation. Evaluating the sum, we have

and

$$[\operatorname{Ca}]^{pre} = \overline{g} e^{-t/\tau} (a+bV_R) \tau_2 [B_{\infty}(1-\mu)+\mu] \\ \times \left\{ (e^{1/(\nu\tau_2)}-1) e^{t_1^{pre}/\tau} \left[\frac{1-\exp\left(\frac{N-1}{\nu\tau}\right)}{1-\exp\left(\frac{1}{\nu\tau}\right)} \right] \\ + \exp\left(\frac{t_N^{pre}}{\tau_N} + \frac{t}{\tau_2}\right) - \exp\left(\frac{t_N^{pre}}{\tau}\right) \right\},$$
(21)

where N and t_N^{pre} are defined as in Eqs. (20b) and (20c).

To calculate the contribution to the calcium transients due to the interaction of the presynaptic and postsynaptic spikes, we substitute the expressions (14) and (17) into Eq. (18b). Let $t_m^{post} \equiv t_n^{pre} + \Delta t$ and $t_M^{post} \equiv t_N^{pre} + \Delta t$. The indexes *m* and *M* will always refer to the postspikes, while *n* and *N* will always refer to the prespikes; for simplicity, we drop the superscripts so that $t_n^{pre} = t_n$ and $t_m^{post} = t_m$. Because the depolarization due to each postspike does not build up, the product of the sums given by expressions (17) and (14) will yield only four terms, due to the interaction of the pairs $\{(n,m-1);(n,m);(N,M-1);(N,M)\}$. These terms are

$$\int_{0}^{t} f(t') V_{B}(t') dt'$$

$$= v [B_{\infty}(1-\mu) + \mu]$$

$$\times \left\{ \sum_{n=2}^{N-1} \int_{t_{n}}^{t_{m}} \exp\left(-\frac{t'-t_{n}}{\tau_{N}} - \frac{t'-t_{m-1}}{\tau_{B}} + \frac{t'}{\tau}\right) dt' \quad (22a)$$

$$+ \sum_{n=1}^{N-1} \int_{t_{m}}^{t_{n+1}} \exp\left(-\frac{t'-t_{n}}{\tau_{N}} - \frac{t'-t_{m}}{\tau_{B}} + \frac{t'}{\tau}\right) dt' \quad (22b)$$

$$+ \int_{t_N}^{t_*} \exp\left(-\frac{t'-t_N}{\tau_N} - \frac{t-t_{M-1}}{\tau_B} + \frac{t'}{\tau}\right) dt'$$
 (22c)

$$+ \delta_{M,N} \int_{t_M}^t \exp\left(-\frac{t'-t_N}{\tau_N} - \frac{t'-t_M}{\tau_B} + \frac{t'}{\tau}\right) dt'\}, \quad (22d)$$

where $t_* = \min(t, t_M)$ and we have used that $B_n = B_N = B_\infty$. Recalling expressions (20a) through (20c), one can easily perform the integrations (22a) through (22d). The remaining sums are finite power series, therefore they converge. Thus, the calcium concentration due to the interaction between presynaptic and postsynaptic spikes will be

$$[Ca]^{+/-} = \overline{g}e^{-t/\tau}bv[B_{\infty}(1-\mu)+\mu]$$

$$\{(22a)+(22b)+(22c)+(22d)\}, \qquad (23)$$

with each of the terms being

$$(22a) = \tau_{3} \left[\exp\left(\frac{\Delta t}{\tau_{2}} - \frac{1}{\nu \tau_{B}}\right) - \exp\left(\frac{\Delta t - 1/\nu}{\tau_{B}}\right) \right]$$

$$\times e^{t_{1}^{pre}/\tau} \left[\frac{1 - \exp\left(\frac{N-1}{\nu \tau}\right)}{1 - \exp\left(\frac{1}{\nu \tau}\right)} \right],$$

$$(22b) = \tau_{3} \left[\exp\left(\frac{\Delta t}{\tau_{B}} + \frac{1}{\nu \tau_{3}}\right) - \exp\left(\frac{\Delta t}{\tau_{2}}\right) \right]$$

$$\times e^{t_{1}^{pre}/\tau} \left[\frac{1 - \exp\left(\frac{N-1}{\nu \tau}\right)}{1 - \exp\left(\frac{1}{\nu \tau}\right)} \right],$$

$$(22c) = \begin{cases} \tau_{3} \left[\exp\left(\frac{\Delta t}{\tau_{2}} - \frac{1}{\nu \tau_{B}}\right) - \exp\left(\frac{\Delta t - 1/\nu}{\tau_{B}}\right) \right] \\ \times \exp\left(\frac{t_{N}^{pre}}{\tau}\right) & \text{if } (t_{N}^{pre} + \Delta t) < t, \end{cases}$$

$$(22c) = \begin{cases} \tau_{3} \left[\exp\left(\frac{t_{N}^{pre}}{\tau_{1}} + \frac{t}{\tau_{3}}\right) - \exp\left(\frac{t_{N}^{pre}}{\tau}\right) \right] \\ \times \exp\left(\frac{\Delta t - 1/\nu}{\tau_{B}}\right) & \text{if } (t_{N}^{pre} + \Delta t) > t, \end{cases}$$

$$(22d) = \begin{cases} \tau_{3} \left[\exp\left(\frac{t_{M}^{post}}{\tau_{1}} + \frac{t}{\tau_{3}}\right) - \exp\left(\frac{t_{M}^{post}}{\tau}\right) \right] \\ \times \exp\left(-\frac{\Delta t}{\tau_{N}}\right) & \text{if } (t_{N}^{pre} + \Delta t) > t, \end{cases}$$

Plots of these expressions are shown in Fig. 5. As expected, the overall calcium concentration is frequency- and time-dependent. There is temporal integration of calcium levels, so that higher-frequency stimulations build up the intracellular calcium concentration [Fig. 5(a)]. In addition, the Δt dependence seen in previous sections [Fig. 5(b)] for single pairs of prespike and postspike is retained. The amount of temporal integration will naturally depend on the specific values of τ , $\tau_{\rm N}$, and $\tau_{\rm B}$, and so will the difference between the pre-post and the post-pre conditions, as analyzed before. For example, slower dynamics would result in more time integration at moderate frequencies. However, given these constants, it is possible to set a threshold of calcium concentration between LTD and LTP (or between noplasticity and LTD) that would correspond to low- and highfrequency STDPs, respectively.

We now extract an expression for the dependence of calcium transients on Δt and on ν in the limit $t \rightarrow \infty$. Let $t = t_N^{pre} + \delta$, where the new variable δ tracks the time since the last presynaptic spike, and define $\gamma_1 = \overline{g}(a+bV_R)\tau_2[B_{\infty}(1-\mu)+\mu]$ and $\gamma_2 = \overline{g}bV_B\tau_3[B_{\infty}(1-\mu)+\mu]$. For $t \rightarrow \infty$ and $\Delta t < \delta$, we have



$$[Ca](\delta) = \gamma_{1}e^{-\delta/\tau} \left\{ e^{\delta/\tau_{2}} + \left[\frac{e^{1/\nu\tau_{2}} - e^{1/\nu\tau}}{e^{1/\nu\tau} - 1} \right] \right\} + \gamma_{2}e^{-\delta/\tau} \\ \times e^{\Delta t/\tau_{B}} \left\{ \left[\frac{e^{1/\nu\tau_{3}} - e^{\Delta t/\tau_{3}}}{e^{1/\nu\tau} - 1} \right] + \exp\left(\frac{-1}{\nu\tau_{B}} + \frac{1}{\nu\tau} \right) \\ \times \left[\frac{e^{\Delta t/\tau_{3}} - 1}{e^{1/\nu\tau} - 1} \right] + \left(e^{\delta/\tau_{3}} - e^{\Delta t/\tau_{3}} \right) \right\}.$$
(24)

And for $\Delta t > \delta$,

$$\begin{split} [Ca](\delta) &= \gamma_1 e^{-\delta/\tau} \Biggl\{ e^{\delta/\tau_2} + \Biggl[\frac{e^{1/\nu\tau_2} - e^{1/\nu\tau}}{e^{1/\nu\tau} - 1} \Biggr] \Biggr\} \\ &+ \gamma_2 e^{-\delta/\tau} e^{\Delta t/\tau_B} \Biggl\{ [e^{\Delta t/\tau_3} - 1] \Biggl[\frac{e^{-1/\nu\tau_B}}{e^{1/\nu\tau} - 1} \Biggr] \\ &+ \Biggl[\frac{e^{1/\nu\tau_3} - e^{\Delta t/\tau_3}}{e^{1/\nu\tau} - 1} \Biggr] + (e^{\delta/\tau_3} - 1) e^{-1/\nu\tau_B} \Biggr\}. \end{split}$$

$$(25)$$

These expressions show how the peaks of the calcium transients depend on the timing and on the frequency of the presynaptic and postsynaptic spikes. As the frequency increases [Fig. 6(a)] the Δt -dependent curves move up due to temporal integration, and the difference between pre-post and post-pre conditions [Fig. 6(b)] becomes smaller.



FIG. 5. (a) Rate-dependence of the calcium transients as derived in expressions (22a) through (22d) for $\Delta t = 10$ ms at 5 Hz (solid line) and 10 Hz (dashed line). (b) Calcium transients at 5 Hz for $\Delta t = +10$ ms (solid line) and $\Delta t = -10$ ms (dotted line). All parameters are as in Fig. 1.

V. STOCHASTIC PROPERTIES OF CALCIUM INFLUX

In previous sections we have calculated the mean calcium transients under various conditions and assumptions. However, synaptic transmission is a random process and trial-bytrial calcium transients deviate from the mean. One source of variability is the stochasticity of the presynaptic neurotransmitter release. A failure of release at a given synapse would result in eliminating the postsynaptic calcium transient at that synapse. Although this source of variability can be significant, its consequences are simple to predict, because the absence of release will result in no synaptic plasticity in the specific spine. Another source of variability is the stochasticity of the postsynaptic opening of NMDA channels. Because the number of NMDAR at each synapse can be small (≈ 10) [30], these fluctuations could be considerable.

In this section, we calculate the variability due to Z NM-DAR in the postsynaptic terminal. We assume the following simple Markov model: the state vector $\mathbf{P}_n(t)$ denotes the probability of being in each one of the *n* possible states and the transition matrix R_n represents transition probabilities from state to state. For a Markov processes, the evolution equation has the form

$$\frac{d\mathbf{P}_n}{dt} = R_n \mathbf{P}_n \,, \tag{26}$$

with the solution

$$\mathbf{P}_{n}(t) = \mathbf{P}_{n}(0) \exp\left(\int_{0}^{t} R_{n} dt'\right), \qquad (27)$$

FIG. 6. (a) Peak calcium transients as a function of the timing between presynaptic and postsynaptic spikes for three different frequencies: 1 Hz (circle), 5 Hz (square), and 10 Hz (triangle). (b) Peak calcium transients as a function of frequency for pre-post (square) and a post-pre (circle) conditions. where $\mathbf{P}_n(0)$ is the initial condition.

In the simplest model the process has only two states, unbound u and open-bound o,

$$k_1G \\ \rightarrow \\ u \quad o, \\ \leftarrow \\ k_{-1} \tag{28}$$

where G is the glutamate concentration, and k_1 and k_{-1} are the forward and backward time constants, respectively.

This model is clearly simplified; typically, a five-state model is used to account for NMDAR kinetics. However, it is sufficient for describing the single exponential decay kinetics we have assumed for the NMDAR since, during decay, G=0 [see Eq. (31), below]. The transition matrix for this model is

$$R_2 = \begin{pmatrix} -k_1 G & k_{-1} \\ k_1 G & -k_{-1} \end{pmatrix},$$
 (29)

where G is the neurotransmitter concentration. We assume that G has a constant, nonzero value for only a limited amount of time. We are primarily interested in the falling phase (G=0) of the NMDAR current.

We denote by p^u and p^o the probabilities of being in the unbound and open states, respectively. Since $p^u + p^o = 1$ the differential equation for p^o reads

$$\dot{p}^{o} = k_{1}G - p^{o}(k_{1}G + k_{-1}).$$
 (30)

For a constant value of G, the solution of this equation is

$$p^{o}(t) = \frac{k_{1}G}{k_{1}G + k_{-1}} + Ce^{-(k_{1}G + k_{-1})t}.$$
 (31)

where the constant C is determined by the initial conditions.

During the falling phase (G=0) the time constant has the value $\tau_{\rm N} = 1/k_{-1}$. The solution takes the form $p^o(t) = \mu e^{-t/\tau_{\rm N}}$, where μ can now be defined more precisely as the open probability at the start of the falling phase. The total calcium current through Z identical NMDAR is $I(t) = \sum_{i=1}^{Z} s_i g \mathcal{H}(V(t))$, where g is the single channel NMDAR conductance, and $s_i = 1$ when receptor is in an open state, and 0 when in closed and unbound states. To simplify the notation we rewrite Eq. (7) as $\mathcal{H} = V_0 + V_1 \Theta(t - t^{post}) e^{-(t-t^{post})/\tau_{\rm B}}$, where $V_0 = a + b V_R$ and $V_1 = bv$.

Calcium concentration is assumed to be governed by Eq. (1), therefore the average calcium concentration will be

$$\langle [Ca](t) \rangle = g e^{-t/\tau} \int_{-\infty}^{t} dt' e^{t'/\tau} \left\langle \sum_{i=1}^{Z} s_i(t') \right\rangle \mathcal{H}(t'),$$
(32)

where $\langle \rangle$ denotes averaging over the probability distribution of $s_i(t)$.

However, $\langle \Sigma s_i(t) \rangle = Z \langle s_i(t) \rangle = Z p^o(t)$, thus the calcium concentration for given Z NMDAR is

$$\langle [Ca](t) \rangle_{Z} = Zg e^{-t/\tau} \int_{-\infty}^{t} dt' e^{t'/\tau} p^{o}(t') \mathcal{H}(t').$$
(33)

Expression (33) is similar to Eq. (2) since $I(t) = Zg\mathcal{H}(t)p^o(t)$. We have calculated this integral in Sec. II for the different cases (pre-only, pre-post, post-pre conditions). These assumptions are equivalent to the simple Markov model assumed here. This result is shown in Eqs. (32) and (33). We note that $\langle [Ca](t) \rangle_Z = Z \langle [Ca](t) \rangle_1$.

The variance for Z NMDAR has the form $\sigma_Z^2(t) = \langle [Ca]^2(t) \rangle_Z - \langle [Ca](t) \rangle_Z^2$. The second moment of the calcium concentration for Z NMDAR is

$$\langle [Ca]^{2}(t) \rangle_{Z} = g^{2} e^{-2t/\tau} \int_{-\infty}^{t} dt' dt'' e^{t'/\tau} e^{t''/\tau} \mathcal{H}(t') \mathcal{H}(t'') \times \left\langle \sum_{i=1}^{Z} s_{i}(t') \sum_{j=1}^{Z} s_{j}(t'') \right\rangle = g^{2} e^{-2t/\tau} \int_{-\infty}^{t} dt' dt'' e^{t'/\tau} \mathcal{H}(t') \mathcal{H}(t'') \times \sum_{i,j=1}^{Z} p_{ij}^{oo}(t',t''),$$
(34)

where $p_{ij}^{oo}(t,t')$ is the joint probability of having channel *i* open at time *t* and channel *j* open at time *t'*. We assume independent identical channels, therefore for $i \neq j$ we have $p_{ij}^{oo}(t,t') = p^{o}(t)p^{o}(t')$, and when i = j, $p_{ii}^{oo}(t,t')$ is defined as $p^{oo}(t,t')$. We therefore have that

$$\langle [Ca]^{2}(t) \rangle_{Z} = g^{2} e^{-2t/\tau} \int_{-\infty}^{t} dt' dt'' e^{t'/\tau} e^{t''/\tau} \mathcal{H}(t') \mathcal{H}(t'') \times [Z(Z-1)p^{o}(t')p^{o}(t'') + Zp^{oo}(t',t'')] = g^{2}Z(Z-1) \langle [Ca](t) \rangle^{2} + g^{2}Ze^{-2t/\tau} \times \int_{-\infty}^{t} dt' dt'' e^{t'/\tau} e^{t''/\tau} \mathcal{H}(t') \mathcal{H}(t'')p^{oo}(t',t'').$$
(35)

Thus the variance of the calcium concentration for Z NM-DAR is

$$\sigma_{Z}^{2}(t) = g^{2} Z e^{-2t/\tau} \int_{-\infty}^{t} dt' dt'' e^{t'/\tau} e^{t''/\tau} \mathcal{H}(t') \mathcal{H}(t'') \times [p^{oo}(t',t'') - p^{o}(t')p^{o}(t'')] = g^{2} Z e^{-2t/\tau} \int_{-\infty}^{t} dt' dt'' \mathcal{H}(t') \mathcal{H}(t'') e^{t'/\tau} e^{t''/\tau} p^{oo}(t',t'') - Z \langle [Ca](t) \rangle^{2}.$$
(36)

We can use the Bayes rule to rewrite $p^{oo}(t,t') = p^{oo}(t|t')p^{o}(t')$. There are two variables which we need to



calculate from the Markov process, $p^{o}(t)$ and $p^{oo}(t|t')$. The solution for $p^{o}(t)$ is given by Eq. (31). We define the time t=0 as the beginning of the falling phase (G=0), or t^{pre} =0. Further, we assume an instantaneous rise time. So for $t < 0, p^{o}(t) = 0$ and at $t = 0 p^{o}(0) = \mu$. Thus for $t > 0 p^{o}(t)$ $=\mu e^{-t/\tau_{\rm N}}.$

Similarly, $p^{oo}(t|t')$ is the probability of being in open state at time t given that the channel was in open state at time t'. We calculate this for the case $t, t' \ge 0$. If t' > t this can be solved in a similar manner to p^{o} , since in this regime it is a stationary Markov process, and must be a function of (t -t'), and at t=t, $p^{oo}(t=t'|t')=1$. During the falling phase there is one directional movement from o to u, since G=0. Therefore, if we know that at time t' the channel is in an open state, it must have been in an open state at any time t< t'. Therefore,

$$p^{oo}(t|t') = \begin{cases} e^{-(t-t')/\tau_{\rm N}}; & t > t' \\ 1; & t \le t'. \end{cases}$$
(37)

Note that this holds for (t,t') > 0, and that by definition at times smaller than zero the receptor is unbound. Hence,

$$p^{oo}(t,t') = \begin{cases} \mu e^{-t/\tau_{\rm N}}; & t > t' \\ \mu e^{-t'/\tau_{\rm N}}; & t < t'. \end{cases}$$
(38)

This expression can now be substituted into Eq. (36), and by using the linear approximation of \mathcal{H} we obtain an expression for $\sigma_Z^2(t)$ that can be computed analytically. To aid in calculation we denote

$$\sigma_{Z}^{2}(t) = Z \mu g^{2} e^{-2t/\tau} (V_{0}^{2} \alpha_{1} + 2V_{0}V_{1}\alpha_{2} + V_{1}^{2}\alpha_{3}) - Z \langle [Ca](t) \rangle^{2},$$
(39)

where

$$\alpha_1 = \int_0^t dt' \int_0^t dt'' p^{oo}(t',t'') e^{t'/\tau} e^{t''/\tau} \Theta(t), \quad (40a)$$

$$\alpha_2 = \int_0^t dt' \int_0^t dt'' e^{-(t'-t^{post})/\tau_{\rm B}} p^{oo}(t',t''), \qquad (40b)$$

$$\alpha_{3} = \int_{0}^{t} dt' \int_{0}^{t} dt'' e^{-(t'-t^{post})/\tau_{\rm N}} e^{(t''-t^{post})/\tau_{\rm B}} p^{oo}(t',t'')$$
$$\times e^{t'/\tau} e^{t''/\tau} \Theta(t'-t^{post}) \Theta(t''-t^{post}). \tag{40c}$$



FIG. 8. Coefficient of variation as a function of Δt for Z = 10and μ =0.5. Note that the variability decreases with increasing values of $\tau_{\rm N}$. Shown are the plots for $\tau_{\rm N}$ =50 ms (circle), $\tau_{\rm N}$ = 75 ms (square), and $\tau_{\rm N}$ = 100 ms (diamond).

300

FIG. 7. Mean calcium transients (solid line) and their standard deviation (dash-dotted line) for different values of Δt , with Z = 10 and μ = 0.5. (a) Δt = 10 ms, (b) $\Delta t = -10$ ms, and (c) Δt = 60 ms. The remaining parameters are as in Fig. 2. Note that the peaks of the calcium transients differ from that of Fig. 2 because μ and Z are different.

In Appendix A we analytically calculate the form of the terms α_1 , α_2 and α_3 . Although the analytical form is complex, the consequences are simple and significant. To quantify the dependence of the relative variability on Δt , we compute the coefficient of variation, $C_V(Z) = \sigma_Z / \langle [Ca] \rangle$ $(\times(t))_{Z}$, where both $[Ca](t)_{Z}$ and $\sigma_{Z}(t)$ are measured at the time t where $\langle [Ca](t) \rangle_Z$ is maximal. As expected, $C_V(Z)$ $=C_V(1)/\sqrt{Z}$. Thus the variability is significant only at relatively low Z. In Fig. 7 we show $\langle [Ca](t) \rangle_Z$ and σ_Z , for Δt = -10, 10, and 60 ms. In all cases both the mean and σ_Z change over time. The peak of $\langle [Ca](t) \rangle_Z$ for $\Delta t = -10$ ms and $\Delta t = 60$ ms are similar. However, the magnitude of the variability, σ_Z , is significantly larger at $\Delta t = 60$ ms than at $\Delta t = -10$ ms.

When $\Delta t < 0$, C_V is low and decreases as $\Delta t \rightarrow 0$ from below. For $\Delta t > 0$, C_V increases as Δt increases (Fig. 8). It is interesting to compare the C_V for values of $\Delta t < 0$ and Δt >0 which have similar peak calcium levels. For example, for $\Delta t = -10$ ms, $C_{V}(10) = 0.34$ and for $\Delta t = 60$ ms, which has a similar peak calcium level, $C_V(10) = 0.51$. Therefore $C_V(Z=10,\Delta t=60)/C_V(Z=10,\Delta t=-10)\approx 1.5.$ Because $C_V(Z) = C_V(1)/\sqrt{Z}$, the relative amplitudes of C_V for varying Δt are independent of Z.

The variability of calcium transients also depends on μ (Fig. 9), decreasing as μ increases, for all Δt . The variable μ is the probability of glutamate binding to postsynaptic receptors given a presynaptic spike, and can be taken to be the



presynaptic probability of release. Thus, as the presynaptic probability of release increases, C_V decreases.

VI. DISCUSSION

Calcium transients due to NMDA currents are believed to play a major role in many forms of synaptic plasticity. We have recently shown that a model based on these transients can account for various different plasticity-induction protocols [16]. In this paper, we computed analytically the calcium dynamics evoked by pairs of presynaptic and postsynaptic spikes under different conditions, so that the variables that control calcium transients and impact synaptic plasticity can be investigated.

We showed that the peak of the calcium transients depends on the Δt : for $\Delta t > 0$, the peak concentration decays with the time constant τ_N of the NMDAR, and for $\Delta t < 0$, it decays with the time constant $au_{\rm B}$ of the BPAP function $V_{R}(t)$. Therefore, if spike-time-dependent plasticity indeed depends on the calcium transients, $\tau_{\rm N}$ and $\tau_{\rm B}$ will determine the width of the pre-post LTP and the post-pre LTD windows, respectively. These results were confirmed by simulations, in which the approximations we used in the calculations were relaxed. The Δt dependence of the peak calcium transient is similar in the more complex and realistic cases where the BPAP dynamics is composed of a fast and a slow component: the difference lies in the $\Delta t < 0$ case, which now depends on the combination of the two time constants. The two-component BPAP allows a sharper transition between the post-before-pre and the pre-before-post STDP windows, which is further enhanced by a fast calcium dynamics. We also showed that, besides the timing of the prespike and postspike, the peak values of the calcium transients also depend on the frequency ν of the presynaptic and postsynaptic conditioning, increasing for greater values of ν . However, at higher frequencies, the dependence on Δt decreases. The amount of calcium built up at a given frequency depends on the system parameters; slower time constants results in more temporal integration. Temporal integration of calcium transients as described here can explain why, in some cases, the induction of STDP is frequency-dependent.

One of the predictions of the unified calcium model is that there exists a pre-before-post LTD for values of Δt greater than those that elicit LTP [16]. This has been shown experimentally by some investigators [20]; others have only placed a small number of data points in this region [8,21]. In all published STDP experiments, there is a large variability in the magnitude and the sign of synaptic plasticity across the different values of Δt . It would therefore be difficult to assess the existence of this type of LTD without a large amount of data. The variability encountered in the experiments indicates that it should be important to examine the variation of the calcium transients, in addition to their mean.

We calculated the variance of the calcium transients and showed that, for a small number Z of NMDAR, this variance can be quite significant. There are indications that the number of NMDAR in the dendrite is indeed small (≈ 10) [30]. Further, we showed that the C_V increases monotonically with Δt , for $\Delta t > 0$. These fluctuations can have significant implications on the outcome of downstream processes dependent on calcium, such as phosphorylation or other secondmessenger cascades upon which plasticity relies.

Previously, several authors have simulated and analyzed dynamical models of synaptic plasticity that can lead to STDP [9,12,14,15,31]. These models typically bypass detailed descriptions of the physiology and biochemistry underlying synaptic plasticity. Recently, biophysical models have been receiving increasing focus [10,11,13,16,32]. These more mechanistic approaches have allowed one to show that various different induction protocols can be accounted for by a single set of assumptions regarding the dependence of plasticity on calcium concentrations [16]. In these models, plasticity is a highly nonlinear function of calcium concentration. Therefore, fluctuations of calcium transients will not only add spread to the resulting LTD and LTP but will also shift their mean (the mean of the solution is not equivalent to the solution given the mean calcium level). For example, in the unified calcium model [16], the sign and magnitude of synaptic plasticity is determined by a saturated U-shaped function of calcium concentration. When calcium falls below a threshold θ_d , no plasticity occurs, when it falls between θ_d and θ_p , $\theta_p > \theta_d$, LTD is induced, and for calcium above θ_p , LTP is induced. Thus, in the LTD region, if the spread of the calcium is of the order of $\theta_p - \theta_d$, occasional LTP can still happen. We showed an explicit example for the cases of Δt = -10 ms and $\Delta t = 60$ ms, where the mean peak calcium concentrations are similar, but the variability for the latter is larger. If this mean calcium is in the LTD range, it is likely that the observed phenomenon is that the LTD is stronger and more robust for $\Delta t = -10$ ms than for $\Delta t = 60$ ms.

This work is based on several simplifying assumptions. This allows us a qualitative understanding of the aspects of calcium dynamics that are crucial for the induction of synaptic plasticity. Incorporation of more realistic features, such as those assumed in the unified calcium model, would probably yield quantitatively more precise results. For example, a double-component NMDAR decay would allow for more calcium build up without sacrificing the temporal asymmetry between pre-post and post-pre conditions. However, we believe that, despite the constraints of analytical tractability, this study can serve as a basis for future models that aim to formalize the biophysical basis of synaptic plasticity.



FIG. 9. Coefficient of variation for Z=10 as a function of μ , for three different values of Δt : $\Delta t = -10$ ms (diamond), $\Delta t = 10$ ms (square), and $\Delta t = 60$ ms (circle). As μ increases, C_V decreases.



FIG. 10. The error estimation for the linear approximation of the \mathcal{H} function. (a) The original \mathcal{H} function (dotted line) and its linear approximation (solid line) in the interval [-70, -10] mV. (b), (c), and (d) The calcium transients elicited by the pre-only, post-pre $(\Delta t = -10 \text{ ms})$, and pre-post ($\Delta t = 10 \text{ ms}$) conditions, respectively, for the full expression (dotted line) and for the linear approximation (solid line).

ACKNOWLEDGMENTS

H.Z.S. would like to thank the members of the IBNS at Brown University as well as M. Tsodyks, D. Golomb, and Y. Kubota for useful discussions and comments, and the ITP at Santa Barbara where some of this work was carried out. This work was partly supported by the Burroughs-Wellcome Fund and the Charles A. Dana Foundation.

APPENDIX A: VARIANCE CALCULATION

Using the definition in Eq. (40a) through Eq. (40c) we obtain

$$\begin{aligned} \alpha_{1} &= \left[\int_{0}^{t} dt' \int_{t'}^{t} dt'' e^{-t''/\tau_{N}} e^{t'/\tau} e^{t''/\tau} \\ &+ \int_{0}^{t} dt' \int_{0}^{t'} dt'' e^{-t''/\tau_{N}} e^{t''/\tau} e^{t''/\tau} \right] \Theta(t) \\ &= \left[\int_{0}^{t} dt' e^{t'/\tau} \tau_{2} (e^{t/\tau_{2}} - e^{t'/\tau_{2}}) \\ &+ \int_{0}^{t} dt' e^{t'/\tau_{2}} \tau (e^{t'/\tau_{1}} - 1) dt' \right] \Theta(t) \\ &= \left[e^{t/\tau_{2}} \tau_{2} \tau (e^{t/\tau} - 1) - \tau_{2} \int_{0}^{t} dt' e^{t'/\tau_{4}} \\ &+ \tau \int_{0}^{t} dt' e^{t'/\tau_{4}} - \tau \int_{0}^{t} dt' e^{t'/\tau_{2}} \right] \Theta(t) \\ &= \left[e^{t/\tau_{4}} (\tau_{2} \tau - \tau_{2} \tau_{4} + \tau \tau_{4}) - 2e^{t/\tau_{2}} \tau_{2} \tau \\ &+ (\tau_{2} \tau_{4} - \tau \tau_{4} + \tau_{2} \tau) \right] \Theta(t), \end{aligned}$$

where
$$\tau_4^{-1} = 2 \tau^{-1} - \tau_N^{-1}$$
.

$$\alpha_{2} = e^{t^{post/\tau_{B}}} [\tau_{5}(\tau - \tau_{2})(e^{t/\tau_{5}} - e^{\tilde{t}/\tau_{5}}) - \tau\tau_{3}(e^{t/\tau_{3}} - e^{\tilde{t}/\tau_{3}}) + \tau_{2}\tau_{6}e^{t/\tau_{2}}(e^{t/\tau_{6}} - e^{\tilde{t}/\tau_{6}})]\Theta(t - \tilde{t}),$$
(A2)

where τ_3 is as defined before, $\tau_5^{-1} = 2 \tau^{-1} - \tau_N^{-1} - \tau_B^{-1}$, $\tau_6^{-1} = \tau^{-1} - \tau_B^{-1}$, and

$$\tilde{t} = \begin{cases} t^{post} & t^{post} > 0\\ 0 & t^{post} \le 0. \end{cases}$$
(A3)

Finally,

$$\alpha_{3} = 2e^{2t^{post}/\tau_{\rm B}}\tau_{6}\tau_{3}[e^{t/\tau_{6}} - e^{t/\tau_{3}}e^{\tilde{t}/\tau_{6}} + e^{\tilde{t}/\tau_{6}}]\Theta(t-\tilde{t}).$$
(A4)

APPENDIX B: NUMERICAL ESTIMATION OF THE ERROR RESULTING FROM THE LINEAR APPROXIMATION OF H

In the simulations, we used the following form for the voltage-dependence of the NMDAR:

$$\mathcal{H}(V) = \frac{V - V_{rev}}{1 + \frac{e^{-0.062V}}{3.57}},$$
(B1)

where $V_{rev} = 130$ mV.

Such functional dependence captures the qualitative nonlinear dependence of the NMDAR on the voltage, although the reversal potential for calcium might be inaccurate at the values close to V_{rev} . In our analysis, we approximated \mathcal{H} as being linear. The fit used is shown in Fig. 10(a). We see that over the relevant range ([-70 0]) this is a relatively good approximation.

In panels (b)–(d) of Fig. 10 we compare the numerically extracted calcium transients (dashed line), using the nonlinear \mathcal{H} and the analytical results obtained for the linear approximation (solid line). At rest [Fig. 10(b)], the linear approximation significantly underestimates the magnitudes of the transients, because this is where the linear fit deviates the most from the true curve. However, in the working regime ([-10, 10] mV), the approximation can be considered adequate.

Let Eq. (B1) be rewritten as $\mathcal{H}(v) = a + bV$ as in Eq. (7). The parameters of linearization are $a = 0.1031/\overline{g}$ and $b = 0.0015/\overline{g}$, where $\overline{g} = -10^{-3}$ is the average NMDA conductance.

- M.F. Bear and R.C. Malenka, Curr. Opin. Neurobiol. 4, 389 (1994).
- [2] J.A. Cummings, R.M. Mulkey, R.A. Nicoll, and R.C. Malenka, Neuron 16, 825 (1996).
- [3] S.N. Yang, Y.G. Tang, and R. Zucker, J. Neurophysiol. 81, 781 (1999).
- [4] S.M. Dudek and M.F. Bear, Proc. Natl. Acad. Sci. U.S.A. 89, 4363 (1992).
- [5] T.V.P. Bliss and G.L. Collingridge, Nature (London) 361, 31 (1993).
- [6] D.E. Feldman, R.A. Nicoll, R.C. Malenka, and J.T. Isaac, Neuron 21, 347 (1998).
- [7] H. Markram, J. Lübke, M. Frotscher, and B. Sakmann, Science 275, 213 (1997).
- [8] G. Bi and M. Poo, J. Neurosci. 18, 10464 (1998).
- [9] W. Senn, H. Markram, and M. Tsodyks, Neural Comput. 13, 35 (2001).
- [10] T. Kitajima and K. Hara, Neural Networks 13, 445 (2000).
- [11] U.R. Karmarkar and D.V. Buonomano, J. Neurophysiol. 88, 507 (2002).
- [12] H.D. Abarbanel, R. Huerta, and M.I. Rabinovich, Proc. Natl. Acad. Sci. U.S.A. 99, 10132 (2002).
- [13] H.Z. Shouval, G.C. Castellani, L.C. Yeung, B.S. Blais, and L. N Cooper, Biol. Cybern. 87, 383 (2002).
- [14] R. Kempter, W. Gerstner, and J.L. van Hemmen, Phys. Rev. E 59, 4498 (1999).
- [15] S. Song, K.D. Miller, and L.F. Abbott, Nat. Neurosci. 3, 919 (2000).

- [16] H.Z. Shouval, M.F. Bear, and L. N Cooper, Proc. Natl. Acad. Sci. U.S.A. 99, 10831 (2002).
- [17] J.A. Lisman, Proc. Natl. Acad. Sci. U.S.A. 86, 9574 (1989).
- [18] A. Artola and W. Singer, Trends Neurosci. 16, 480 (1993).
- [19] M.F. Bear, L. N Cooper, and F.F. Ebner, Science **237**, 42 (1987).
- [20] M. Nishiyama, K. Hong, K. Mikoshiba, M.M. Poo, and K. Kato, Nature (London) 408, 584 (2000).
- [21] P.J. Sjöström, G.G. Turrigiano, and S.B. Nelson, Neuron 32, 1149 (2001).
- [22] J.C. Magee and D. Johnston, Science 275, 209 (1997).
- [23] H. Markram, P.J. Helm, and B. Sakmann, J. Physiol. (London) 485, 1 (1995).
- [24] B.L. Sabatini, T.G. Oerthner, and K. Svoboda, Neuron 33, 439 (2002).
- [25] C.E. Jahr and C.F. Stevens, J. Neurosci. 10, 3178 (1990).
- [26] G. Carmignoto and S. Vicini, Science 258, 1007 (1992).
- [27] D.E. Feldman, Neuron 27, 45 (2000).
- [28] M.E. Larkum, J.J. Zhu, and B. Sakmann, J. Physiol. (London) 533, 447 (2001).
- [29] V.N. Murthy, T.J. Sejnowski, and C.F. Stevens, Proc. Natl. Acad. Sci. U.S.A. 97, 901 (2000).
- [30] C. Racca, F.A. Stephenson, P. Streit, J.D.B. Roberts, and P. Somogyi, J. Neurosci. 20, 2512 (2000).
- [31] N. Levy, D. Horn, I. Meilijson, and E. Ruppin, Neural Networks 14, 815 (2001).
- [32] G.C. Castellani, E.M. Quinlan, L. N Cooper, and H.Z. Shouval, Proc. Natl. Acad. Sci. U.S.A. 98, 12772 (2001).